No Electron-Donating Substituent Effect on the Singlet Excited State Formation from the 5-(5-Aryl-2-pyrazinylamino)-1,2,4-trioxanes in Dimethyl Sulfoxide Triggered by Potassium t-Butoxide

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5-(5-Phenyl-2-pyrazinylamino)-1,2,4-trioxane derivatives having a substituent on the 4 position of the phenyl group gave chemiluminescence in dimethyl sulfoxide by addition of potassium t-butoxide. No electron-donating substituent effect was observed on the yield of singlet excited state formation. This may be explained by the negative charge formed on the nitrogen atom next to the transient dioxetane moiety, which may produce singlet excited-state molecule by the intramolecular CIEEL mechanism.

Simple, isolable dioxetanes such as tetramethyl-1,2dioxetane are relatively stable at room temperature and afford predominantly triplet excited-state molecules on thermolysis,1) whereas singlet excited products were efficiently produced in the case of bioluminescent systems such as firefly and Cypridina²⁾ (Scheme In the latter cases, the transient 1,2-dioxetane intermediate produced must be extremely unstable³⁾ so as in the case of efficiently chemiluminescent compounds. The high efficiency of singlet excited state production and the instability of the intermediate dioxetanes in bioluminescence and efficient chemiluminescence may come from the conjugation of an electron-donating (easily oxidizable) and highly fluorescent chromophore with the excited-state carbonyl group to be formed.4)

Koo et al.⁵⁾ explained these effects in terms of the chemically initiated intramolecular electron-exchange luminescence (intramolecular CIEEL) mechanism. In the firefly case, for example (Scheme 2), electron transfer from the easily oxidizable phenoxide anion to

the highly energized dioxetanone moiety to cause a heterolytic O-O bond fission followed by rapid decarboxylation with electron exchange generates an excited state product (emitter). Thus, the phenoxide anion formation would accelerate strongly the dioxetane decomposition⁶⁾ and singlet state formation.

This is further supported by the observation of Schaap and Gagnon⁷⁾ in chemiluminescence of some dioxetanes. Dioxetane 7b having a p-hydroxyphenyl substituent is fairly stable at room temperature in o-xylene (tau 1/2 at 25 °C, 57 h), but addition of t-BuOK/18-crown-6 dramatically accelerates the rate of decomposition (4.4×106 times) of 7b with produc-

R₁=3-indolyl R₂=CH₂CH₂CH₂NHC(NH)NH₂ R₃=2-methylpropyl

Scheme 1. Mechanism of Cypridina and firefly bioluminescence.

Scheme 2. Schematic presentation of intramolecular CIEEL mechanism.

tion of singlet excited molecules ca. 160 times more than that formed from the neutral molecules. An amino group also has a strongly electron-donating ability and hence is expected to have the effect similar to $-O^{-}$.48-10)

Coelenterazine (*Oplophorus* luciferin) (**8**)²⁾ which is a luminescent chromophore of a photoprotein, aequorin, has a *p*-hydroxyphenyl substituent on the 3,7-dihydroimidazo[1,2-a]pyrazin-3-one chromophore, which is transformed to a dioxetane intermediate during bioluminescent oxidation (see Scheme 1). Shimomura et al.¹¹⁾ previously suggested that the phenolic hydroxyl group must be dissociated in the photoprotein. It might indicate that the phenoxide anion, as in the firefly luciferin case, contributes to the quantum yield of singlet excited state formation, but very recently Shimomura et al.¹²⁾ also suggested that such an anion formation in aequorin would not be important. To see whether this dissociation of the phenolic hydroxyl group is necessary or not to produce a

high-quantum-yield luminescence similar to the case of firefly bioluminescence, we have examined the trioxane **9e** in comparison with those having other substituents, since such the trioxanes **9** have been found to give strong chemiluminescence with base^{13,14)} and are regarded as a luminescent system similar to coelenterazine (**8**) and *Cypridina* luciferin (**1**) (Scheme 3).¹⁵⁾

As shown in Table 1, the p-hydroxyphenyl-substituted trioxane **9e** gave a slightly higher chemiluminescence quantum yield than the phenyl or methoxyphenyl trioxane, **9a** or **9c**, but quantum yield of singlet excited-state molecules was rather decreased since the fluorescence quantum yield of the product **10e** was better than that of others. In this case the phenolic hydroxyl group was dissociated to form phenoxide anion, which has almost no effect on the formation of the singlet excited molecule. **4-Dimethylamino-**phenyl and **3-indolyl** derivatives, **9f** and **9g**, behave

Table 1. Chemiluminescence Properties of the Trioxanes in DMSO Containing *t*-BuOK

Compound	$d \phi_{CL} \times 10^2$	$\phi_F \times 10^2$	$\phi_s^* \times 10^2$	λ_{max}/nm	Reference
9a	0.10	15	0.67	450	16
9 b	0.049	8.1	0.60	465	16
9c	0.14	21	0.67	454	16
9 d	0.13	20	0.65	454	16
9e	0.21	63	0.33	545	This work
9f	0.13	50	0.26	471	This work
9 g	0.22	52	0.42	545	16

 ϕ_{CL} =quantum efficiency of chemiluminescence. ϕ_F =quantum efficiency of fluorescence of the corresponding amide anion 10. ϕ_s^* =quantum efficiency of excited singlet state formation calculated from an equation $\phi_{\text{CL}} = \phi_s^* \times \phi_F$.

- (9a) X=R=H
- (9b) X=Br R=H
- (9c) X=CH₃O R=H
- (9d) X=CH₃O R=benzyl
- (9e) X=OH R=benzyl
- (9f) X=NMe₂ R=benzyl

Scheme 3. Chemiluminescence reaction of the 5-amino-1,2,4-trioxanes.

similarly. The difference in the behavior between the Schaap's compound 7 and the present case may be explained as follows. In the present case, cyclization of the intermediate hydroperoxide anion forms the dioxetane having a negative charge on the nitrogen atom next to the dioxetane moiety. The negative charge donates enough electrons to the dioxetane moiety to produce singlet excited state molecule 10 so

that no other electron-donating moiety would be necessary. On the other hand, firefly luciferin (4) and the Schaap's compound 7a without a phenoxide anion on the molecule have insufficient electron-donating ability to the intermediate dioxetane moiety so that only triplet state produces predominantly. Thus, ionization of the phenolic hydroxyl group in the firefly luciferin (4) and 7b dramatically changes the yield of singlet excited state molecules, whereas no such a change is observed in the present case. This explanation may also be extended to the luminescence of *Cypridina* luciferin (1), since the initially produced excited molecule was proven to be anion of the oxyluciferin (3).¹⁶⁾

Experimental

All melting points were measured on a Mitamura Riken mp aparatus and uncorrected. ¹H NMR spectra were recorded on a JNM-FX200 spectrometer. Chemical shifts are given in ppm from internal TMS and coupling constants in Hz. IR spectra were taken on a JASCO IR-700 infrared spectrometer. UV spectra were obtained on a Hitachi 228 spectrometer. Mass spectra were measured on a JEOL JMS-DX300 instrument. Chemiluminescence and fluorescence spectra were recorded on an Otsuka Electronics MCPD-110A and JASCO FP-770 spectrometers, respectively. Dimethyl sulfoxide was distilled from calcium hydride under reduced pressure. The other solvents were of reagent grade.

5-(4-Dimethylaminophenyl)-3-benzyl-2-aminopyrazine (11f): To a solution of p-dimethylaminophenylglyoxal oxime¹⁷ (3.0 g) and 2-amino-3-phenylpropionitrile (3.0 g) in pyridine (50 ml) was added TiCl₄ (2.0 ml) dropwise under argon at 0 °C. After being allowed to stand for 10 min, the mixture was heated at 50 °C for 30 min. To this reaction mixture was added slowly 10% aq NaHCO₃ at 0 °C to neutralize it and the resulting mixture was filtered through Celite

bed. The filtrate was diluted with water and extracted three times with ethyl acetate. The organic layer was washed with water and sat. NaCl solution, dried over anhydrous Na₂SO₄, and evaporated to dryness. The residue was crystallized from ethyl acetate to give the N-oxide of 11f (2.8 g). The N-oxide was dissolved in MeOH (100 ml) and CH₂Cl₂ (20 ml) and reduced with Raney Ni (W2) at 50 °C under hydrogen atmosphere for 5 h with stirring, and the mixture was filtered. The filtrate was concentrated to a small volume and diluted with diethyl ether to give the aminopyrazine 11f (2.5 g, 52%); slightly brownish needles (from methanol), mp 175—176°C; IR (KBr) cm⁻¹ 3478, 3294, 3134, 1610, 1463; UV (MeOH) λ_{max}/nm (ϵ) 304 (24800), 366 (10500); MS m/z 304 (M⁺); ¹H NMR (CDCl₃) δ (J) 3.00 (6H, s), 4.30 (2H, s), 4.40 (2H, br. s), 6.80 (2H, d, 9.0), 7.30 (5H, s), 7.84 (2H, d, 9.0), 8.30 (1H, s). Calcd for C₁₉H₂₀N₄: C, 74.97; H, 6.62; N, 18.41%. Found: C, 75.00; H, 6.70; N, 18.63%.

5-[5-(4-Dimethylaminophenyl)-3-benzyl-2-pyrazinylamino]-3-isopropyl-6,6-dimethyl-1,2,4-trioxane (9f): A solution of the aminopyrazine 11f (400 mg) and isobutyraldehyde (5.0 ml) in hexane (10 ml) was heated at 90 °C for 3 h and then stirred at 20 °C for 3 days with a CaCl₂ tube. The mixture was poured in 5% aq NaHCO3 and extracted twice with ethyl acetate. The organic layer was dried over anhydrous Na₂SO₄ and evaporated. The residue was chromatographed on a silica-gel column with ethyl acetate-hexane (1:20) to give a mixture of two diastereomers of 9f (350 mg, 58%), which was crystallized from hexane. Recrystallizations of the mixture from hexane gave a main isomer of 9f as slightly brownish needles, mp 129—130°C; IR (KBr) cm⁻¹ 1611, 1499; UV (MeOH) λ_{max}/nm (ϵ) 306 (23900), 362 (12700); MS m/z 462 (M⁺); ¹H NMR (CDCl₃) δ (J) 0.80 (3H, s), 0.90 (3H, d, 7), 0.91 (3H, s, 7), 1.12 (3H, s) 1.79 (1H, m), 3.00 (6H, s), 4.13 (1H, d, 16), 4.30 (1H, d, 16), 4.42 (1H, br. d, 10), 5.12 (1H, d, 5.0), 5.58 (1H, d, 10), 6.84 (2H, br. d, 9.0), 7.25 (5H, m), 7.86 (2H, d, 9.0), 8.40 (1H, s). Calcd for C₂₇N₃₄O₃N₄: C, 70.10; H, 7.41; N, 12.11%. Found: C, 70.23; H, 7.46; N, 12.39%.

5-[5-(4-Hydroxyphenyl)-3-benzyl-2-pyrazinylamino]-3isopropyl-6,6-dimethyl-1,2,4-trioxane (9e): The trioxane 9e was prepared from the aminopyrazine 11e¹⁸⁾ (170 mg), isobutyraldehyde (5.0 ml) and hexane (20 ml) according to the procedure described in the preparation of 9f. After chromatography was obtained 9e (180 mg, 70%) as a 4:1 mixture of diastereomers; slightly yellowish amorphous solid (from ether-hexane); mp 65-67°C; IR (KBr) cm⁻¹ 3430, 1498; UV (MeOH) λ_{max} nm (ϵ) 280 (22200), 346 (10500); MS m/z 435 (M⁺). Calcd for C₂₅N₂₉O₄N₃: C, 68.94; N, 6.71; N, 9.65%. Found: C, 68.89; N, 6.84; N, 9.58%. ¹H NMR $(CDCl_3) \delta (J)$: major isomer $(20 \, ^{\circ}C) 0.80 (3H, s), 0.90 (6H, d, d)$ 6.8), 1.12 (3H, s), 1.75 (1H, m), 4.14 (1H, d, 16), 4.30 (1H, d, 16), 4.50 (1H, d, 10), 5.12 (1H, d, 4.5), 5.58 (1H, d, 10), 5.80 (1H, br. s), 6.90 (2H, d, 8.7), 7.20—7.40 (5H, m), 7.80 (2H, d, 8.7), 8.38 (1H, s); minor isomer (60 °C): 0.77 (3H, d, 7), 0.78 (3H, d, 7), 0.90 (3H, s),1.60 (3H, s) 1.7 (1H, m), 4.14 (1H, d, 15), 4.29 (1H, d, 15), 4.40, (1H, d, 5.0) 5.42 (1H, d, 9.0), 5.58 (1H, d, 9.0), 6.90 (2H, d, 9.0), 7.4 (5H, m), 7.86 (2H, d, 9.0), 8.41 (1H, s).

5-(4-Dimethylaminophenyl)-3-benzyl-2-(formylamino)-pyrazine (10f): To a solution of the aminopyrazine 11f (50 mg) in pyridine (0.8 ml) was added acetic formic anhydride (0.2 ml) dropwise at 0 °C. After being stirred further 2 h, the mixture was evaporated to dryness to give the amide 10f

(45 mg) as yellow needles from methanol; mp 200—202 °C; IR (KBr) 1681 cm⁻¹; UV (MeOH) $\lambda_{\rm max}/{\rm nm}$ (ϵ) 328 (996), 370 (11400); MS m/z 332 (M⁺); ¹H NMR (CDCl₃–CD₃OD, 80 °C) δ (J) 3.00 (6H, s), 4.24 (2H, s), 6.80 (2H, d, 9.5), 7.25 (5H, s), 7.87 (2H, d, 9.5), 8.50 (1H, s), 9.04 (1H, br.s). Calcd for C₂₀N₂₀ON₄: C, 72.27; N, 6.07; N, 16.86%. Found: C, 72.23; N, 6.05; N, 17.03%.

5-(4-Hydroxyphenyl)-3-benzyl-2-formylaminopyrazine (10e): To a solution of the aminopyrazine **11e** (50 mg) in pyridine (0.8 ml) was added acetic formic anhydride (0.2 ml) at 0 °C. After being stirred further 2 h, the reaction mixture was evaporated to dryness and the residue purified by silica gel TLC to give **10e** (20 mg, 36%) as colorless needles from methanol; mp 208—210 °C; IR (KBr) cm⁻¹ 3368, 1674; UV (MeOH) $\lambda_{\text{max}}/\text{nm}$ (ε) 277 (18000), 294 (15800), 341 (15600); MS m/z 305 (M⁺); ¹H NMR (CDCl₃-acetone- d_6) δ (J) 4.30 (2H, s), 6.96 (2H, d, 8.0), 7.28 (5H, s), 7.88 (2H, d, 8.0), 8.20 (1H, br.d, disappeared with D₂O), 8.30 (1H, s, diappeared with D₂O), 8.52 (1H, s), 9.3 (1H, br.s). Calcd for C₁₈N₁₅O₂N₃: C, 70.80; H, 4.95; N, 13.76%. Found: C, 70.83; H, 4.95; N, 13.95%.

Chemiluminescence and Fluorescence Measurements. To a solution of a trioxane (concn 1×10^{-5} mol 1^{-1}) in dimethyl sulfoxide (2 ml) was added a solution of t-BuOK (0.25 mol 1^{-1}) in t-BuOH (0.1 ml) at 25 °C. The resulting light emission was recorded with a luminometer. Quantum yields of chemiluminescence and fluorescence were determined as described previously.⁴)

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References

- 1) W. H. Richardson, J. H. Anderegg, M. E. Price, and R. Crawford, J. Org. Chem., 43, 4045 (1978) and references cited therein
- 2) (Review) T. Goto, "Marine Natural Products," ed by P. Scheuer, Academic Press, Inc., New York (1980), pp. 179—222.

- 3) J. E. Lloyd, "Bioluminescence in Action," ed by P. J. Herring, Academic Press, New York (1978), p. 264.
- 4) T. Goto and H. Nakamura, J. Chem. Soc., Chem. Commun., 1978, 781; H. Nakamura and T. Goto, Photochem. Photobiol., 30, 27 (1979).
- 5) J.-Y. Koo, S. P. Schmidt, and G. B. Schuster, *Proc. Natl. Acad. Sci., USA*, **75**, 30 (1978); J.-Y. Koo and G. B. Schuster, *J. Am. Chem. Soc.*, **99**, 5403 (1977). See also F. McCapra, *J. Chem. Soc.*, *Chem. Commun.*, **1977**, 946.
- 6) E. H. White, H. Worther, H. H. Seliger, and W. D. McElroy, J. Am. Chem. Soc., **88**, 2015 (1966).
- 7) A. P. Schaap and S. D. Gagnon, *J. Am. Chem. Soc.*, **104**, 3504 (1982); see also A. P. Schaap, S. D. Gagnon, and K. A. Zaklika, *Tetrahedron Lett.*, **23**, 2943 (1982); K. A. Zaklika, T. Kissel, A. L. Thayer, P. A. Burns, and A. P. Schaap, *Photochem. Photobiol.*, **30**, 35 (1979).
- 8) C. Lee and L. A. Singer, *J. Am. Chem. Soc.*, **102**, 3823 (1980); K. A. Zaklika, A. L. Thayer, and A. P. Schaap, *ibid.*, **100**, 4916 (1978).
- 9) E. H. White, H. Worther, G. F. Field, and W. D. McElroy, J. Org. Chem., **30**, 2344 (1965).
- 10) K. A. Zaklika, T. Kissel, A. L. Thayer, P. A. Burns, and A. P. Schaap, *Photochem. Photobiol.*, 30, 35 (1979).
- 11) O. Shimomura, F. H. Johnson, and H. Morise, *Biochemistry*, **13**, 3278 (1974).
- 12) O. Shimomura, B. Musick, and Y. Kishi, *Biochem. J.*, **251**, 405 (1988).
- 13) T. Goto and H. Nakamura, Tetrahedron Lett., 1976, 4627.
- 14) H. Nakamura and T. Goto, Bull. Chem. Soc. Jpn., 61, 3776 (1988).
- 15) T. Goto, "New Synthetic Methodology and Functionally Interesting Compounds," ed by Z. Yoshida, Kodansha Ltd., Tokyo (1986), pp. 285—306.
- 16) T. Goto, S. Inoue, K. Sugiura, K. Nishikawa, M. Isobe, and Y. Abe, *Tetrahedron Lett.*, **1968**, 4035.
- 17) J. J. Norman, R. M. Heggie, and J. B. Larose, *Can. J. Chem.*, **40**, 1547 (1962).
- 18) Y. Kishi, H. Tanino, and T. Goto, *Tetrahedron Lett.*, **1972**, 2747.